101.429-12 – including *Taq* polymerase, IFU-01
101.429-12u – without *Taq* polymerase, IFU-02

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Lot No.: 38V **Lot-specific Information** **www.olerup-ssp.com**

Olerup SSP® HLA-A*30

Product number:	**101.429-12 – including *Taq* polymerase** **101.429-12u – without *Taq* polymerase**
Lot number:	**38V**
Expiry date:	**2016-August-01**
Number of tests:	**12**
Number of wells per test:	**31+1**
Storage - pre-aliquoted primers:	**dark at -20°C**
- PCR Master Mix:	**-20°C**
- Adhesive PCR seals	**RT**
- Product Insert	**RT**

This Product Description is only valid for Lot No. 38V.

CHANGES COMPARED TO THE PREVIOUS *OLERUP* SSP® HLA-A*30 LOT (21R)

The HLA-A*30 kit is updated for new alleles to enable separation of:

- Confirmed[1] alleles as listed in the IMGT/HLA database
- Polymorphisms in exons outside of the region encoding the peptide binding domain
- Null and Alternatively expressed alleles

A well containing Negative Control primer pairs has been added.

The format of the Product Insert and Worksheet have been changed.

Two wells have been added to the HLA-A*30 kit, wells **31 to 32**.

[1]As described in section Uniquely Identified Alleles.

The HLA-A*30 specificity and interpretation tables have been updated for the HLA-A alleles described since the previous *Olerup* SSP® HLA-A*30 lot was made **(Lot No. 21R)**.

The primers of the wells detailed below have been exchanged, added or modified compared to the previous lot.

Well	5'-primer	3'-primer	rationale
18	-	Added	3'-primer added for the A*30:71 allele.
19	Added	Added	Primer pair added for the A*30:77 allele, 3'-primer added for the A*30:70N allele.
22	-	Added	3'-primer added for the A*30:73N allele.
24	-	Added	3'-primer added for the A*30:78N allele.
25	Modified	-	Modified 5'-primer for improved HLA-specific amplification.
27	Added	Added	Primer pair added for the A*30:56 allele.
29	-	Added	3'-primer added for the A*30:73N allele.
31	New	New	New primer pair for the A*30:76N allele.
32			Negative Control.

Change in revision R01 compared to R00:
1. Primer mix 24 does not amplify the A*30:78N allele. This has been corrected in the Specificity and Interpretation tables. Thus, this lot of the HLA-A*30 subtyping kit cannot distinguish the A*30:01:01-30:01:07 alleles and the A*30:78N allele.

Well **32** contains Negative Control primer pairs, that will amplify more than 95% of the *Olerup* SSP® HLA Class I, DRB, DQB1 and DPB1 amplicons as well as amplicons generated by a control primer pair.

PCR product sizes range from 75 to 430 base pairs.
The PCR product generated by the control primer pair is 430 base pairs.

Length of PCR product	**105**	**200**	**105**	**80**	**75**	**80**
5'-primer[1]	**164**	**340**	**440**	**45**	**45**	**43**
	5'-CAC3'	**5'-Agg3'**	**5'-TTA3'**	**5'-Tgg3'**	**5'-Tgg3'**	**5'-Tgg3'**
3'-primer[2]	**231**	**2nd I**	**507**	**59**	**58**	**57**
	5'-TgC3'	**5'-AAA3'**	**5'-TTg3'**	**5'-CTC3'**	**5'-ggC3'**	**5'-CTC3'**
A*	+	+	+			
B*	+	+	+			
C*	+	+	+			
DRB1				+	+	
DRB3				+	+	
DRB5				+		
DQB1					+	
DPB1						+

[1]The nucleotide position for HLA class I genes and the codon for HLA class II genes, in the 2nd or 3rd exon, matching the specificity-determining 3'-end of the primer is given. Nucleotide and codon numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

[2]The nucleotide position for HLA class I genes and the codon for HLA class II genes, in the 2nd or 3rd exon or the 2nd intron, matching the specificity-determining 3'-end of the primer is given in the anti-sense direction. Nucleotide and codon numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

101.429-12 – including *Taq* polymerase, IFU-01
101.429-12u – without *Taq* polymerase, IFU-02

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Lot No.: 38V **Lot-specific Information** **www.olerup-ssp.com**

PRODUCT DESCRIPTION

HLA-A*30 SSP subtyping

CONTENT

The primer set contains 5'- and 3'-primers for identifying the A*30:01 to A*30:78N alleles.

PLATE LAYOUT

Each test consists of 32 PCR reactions in a 32 well cut PCR plate

1	**2**	**3**	**4**	**5**	**6**	**7**	**8**
9	**10**	**11**	**12**	**13**	**14**	**15**	**16**
17	**18**	**19**	**20**	**21**	**22**	**23**	**24**
25	**26**	**27**	**28**	**29**	**30**	**31**	**NC**

The 32 well cut PCR plate is marked with 'HLA-A*30' in silver/gray ink.
Well No. 1 is marked with '38V'.
Wells 1 to 31 – HLA-A*30 high resolution primers.
Well 32 – Negative Control (NC).

A faint row of numbers is seen between wells 1 and 2 or wells 7 and 8 of the PCR trays. These stem from the manufacture of the trays, and should be disregarded.
The PCR plates are covered with a PCR-compatible foil.
Please note: When removing each 32 well PCR plate, make sure that the remaining plates stay covered. Use a scalpel or a similar instrument to carefully cut the foil between the plates.

INTERPRETATION

Due to the sharing of sequence motifs between HLA-A alleles non-HLA-A*30 alleles will be amplified by primer mixes 1, 3 to 5, 7, 9 to 12, 17, 19, 20, 28 and 29.
For further details see Specificity Table.

UNIQUELY IDENTIFIED ALLELES

All the HLA-A*30 alleles, i.e. **A*30:01 to A*30:78N alleles**, recognized by the HLA Nomenclature Committee in October 2013[1,2] will be amplified by the primers in the HLA-A*30 subtyping kit.

The HLA-A*30 kit enables separation of the confirmed HLA-A*30 alleles as listed in the IMGT/HLA database. An HLA allele is listed as confirmed by IMGT/HLA if it has been sequenced by more than a single laboratory or from multiple sources. Current allele confirmation status for HLA-A*30 alleles is listed below.

The HLA-A*30 kit also enables identification of polymorphisms in exons outside of the region encoding the peptide binding domain and of null and alternatively expressed alleles

The HLA-A*30 kit cannot distinguish silent mutations in the A*30:01:01- A*30:01:07 alleles, the 30:02:01-30:02:04 and 30:02:06-30:02:12, the A*30:04:01-30:04:02 or the A*30:11:01-30:11:02 alleles.

[1]HLA-A alleles listed on the IMGT/HLA web page 2013-October-11, release 3.14.0, www.ebi.ac.uk/imgt/hla.
[2]Alleles that have been deleted from or renamed in the official WHO HLA Nomenclature up to and including the last IMGT/HLA database release can be retrieved from web page http://hla.alleles.org/alleles/deleted.html.

ALLELE CONFIRMATION STATUS

Allele	Status[1]	Allele	Status[1]	Allele	Status[1]	Allele	Status[1]
A*30:01:01	**Confirmed**	A*30:13	Unconfirmed	A*30:44	Unconfirmed	A*30:74	Unconfirmed
A*30:01:02	Unconfirmed	A*30:14L	Unconfirmed	**A*30:45**	**Confirmed**	A*30:75	Unconfirmed
A*30:01:03	Unconfirmed	A*30:15	Unconfirmed	A*30:46	Unconfirmed	A*30:76N	Unconfirmed
A*30:01:04	**Confirmed**	**A*30:16**	**Confirmed**	A*30:47	Unconfirmed	A*30:77	Unconfirmed
A*30:01:05	Unconfirmed	**A*30:17**	**Confirmed**	A*30:48	Unconfirmed	A*30:78N	Unconfirmed
A*30:01:06	Unconfirmed	A*30:18	Unconfirmed	A*30:49	Unconfirmed		
A*30:01:07	Unconfirmed	A*30:19	Unconfirmed	A*30:50	Unconfirmed		
A*30:02:01	**Confirmed**	**A*30:20**	**Confirmed**	**A*30:51**	**Confirmed**		
A*30:02:02	Unconfirmed	A*30:22	Unconfirmed	A*30:52	Unconfirmed		
A*30:02:03	**Confirmed**	A*30:23	Unconfirmed	A*30:53	Unconfirmed		
A*30:02:04	**Confirmed**	A*30:24	Unconfirmed	A*30:54	Unconfirmed		
A*30:02:05	**Confirmed**	A*30:25	Unconfirmed	A*30:55	Unconfirmed		
A*30:02:06	**Confirmed**	A*30:26	Unconfirmed	**A*30:56**	**Confirmed**		
A*30:02:07	Unconfirmed	A*30:27N	Unconfirmed	A*30:57	Unconfirmed		
A*30:02:08	Unconfirmed	A*30:28	Unconfirmed	A*30:58	Unconfirmed		
A*30:02:09	Unconfirmed	**A*30:29**	**Confirmed**	**A*30:59N**	**Confirmed**		
A*30:02:10	Unconfirmed	**A*30:30**	**Confirmed**	A*30:60	Unconfirmed		
A*30:02:11	Unconfirmed	**A*30:31**	**Confirmed**	**A*30:61**	**Confirmed**		
A*30:02:12	Unconfirmed	**A*30:32**	**Confirmed**	A*30:62	Unconfirmed		
A*30:03	**Confirmed**	A*30:33	Unconfirmed	A*30:63	Unconfirmed		
A*30:04:01	**Confirmed**	A*30:34	Unconfirmed	A*30:64	Unconfirmed		
A*30:04:02	**Confirmed**	A*30:35	Unconfirmed	A*30:65	Unconfirmed		
A*30:06	Unconfirmed	**A*30:36**	**Confirmed**	A*30:66	Unconfirmed		
A*30:07	**Confirmed**	A*30:37	Unconfirmed	A*30:67	Unconfirmed		
A*30:08	**Confirmed**	A*30:38	Unconfirmed	A*30:68	Unconfirmed		
A*30:09	**Confirmed**	A*30:39	Unconfirmed	A*30:69	Unconfirmed		
A*30:10	**Confirmed**	**A*30:40**	**Confirmed**	A*30:70N	Unconfirmed		
A*30:11:01	**Confirmed**	A*30:41	Unconfirmed	**A*30:71**	**Confirmed**		
A*30:11:02	Unconfirmed	**A*30:42**	**Confirmed**	A*30:72	Unconfirmed		
A*30:12	Unconfirmed	A*30:43	Unconfirmed	A*30:73N	Unconfirmed		

[1]Allele status "confirmed" or "unconfirmed" as listed on the IMGT/HLA web page 2013-October-11, release 3.14.0, www.ebi.ac.uk/imgt/hla.

RESOLUTION IN HOMO- AND HETEROZYGOTES

Results file with resolution in HLA-A*30 homo- and heterozygotes is available upon request.

SPECIFICITY TABLE

HLA-A*30 SSP subtyping

Specificities and sizes of the PCR products of the 31+1 primer mixes used for HLA-A*30 SSP subtyping

Primer Mix	Size of spec. PCR product[1]	Size of control band[2]	Amplified HLA-A*30 alleles[3]	Other amplified HLA-A alleles[4]
1	165 bp	**800 bp**	*30:01:01-30:02:04, 30:02:06-30:03, 30:07-30:16, 30:18-30:20, 30:22-30:25, 30:27N-30:28, 30:31-30:45, 30:47-30:76N, 30:78N	*02:52, 03:43, 23:46
2	205 bp	**800 bp**	*30:01:01-30:01:07, 30:08, 30:11:01-30:11:02, 30:14L-30:16, 30:18-30:20, 30:23-30:24, 30:26, 30:30-30:31, 30:35-30:44, 30:48-30:49, 30:53-30:56, 30:58-30:60, 30:62-30:63, 30:65, 30:71-30:75, 30:78N	
3[8]	210 bp	**800 bp**	*30:02:01-30:03, 30:07, 30:10, 30:12-30:13, 30:22, 30:25, 30:27N, 30:32-30:34, 30:45, 30:50-30:51, 30:57, 30:61, 30:64, 30:66-30:70N, 30:76N	*03:05:02, 11:24:01, 34:02:02, 80:01:01:01-80:03
4[6]	155 bp	1070 bp	*30:03, 30:11:01-30:11:02, 30:71	*01:02, 01:20
5	150 bp 245 bp	1070 bp	*30:04:01-30:04:02, 30:06, 30:17, 30:29, 30:46, 30:77 *30:19	*02:52, 03:82, 24:66, 68:06
6[5]	80 bp 185 bp	1070 bp	*30:06 *30:07	
7	200 bp	1070 bp	*30:08, 30:28	*02:185[W]
8[5,8]	85 bp	**800 bp**	*30:10	
9	180 bp	1070 bp	*30:01:01-30:02:12, 30:04:01-30:04:02, 30:06-30:07, 30:09-30:10, 30:12-30:20, 30:23-30:54, 30:56-30:70N, 30:72-30:78N	*24:124
10	150 bp	1070 bp	*30:12, 30:18, 30:55	*01:07, 02:185, 24:124, 26:19, 29:14, 31:01:02-31:01:06, 31:01:08-31:07, 31:09-31:43, 31:45-31:79
11	215 bp	**800 bp**	*30:01:01-30:04:02, 30:06, 30:09-30:20, 30:23-30:30, 30:32-30:54, 30:56-30:59N, 30:61-30:78N	*01:02[W], 01:20[W], 03:72, 11:88, 23:09[W], 23:51, 24:24, 24:67, 24:129[W], 24:145, 24:156, 24:191, 26:16, 29:37, 32:07, 68:45
12[6]	190 bp	1070 bp	*30:09, 30:35	*03:42, 03:131, 03:133, 31:03-31:04, 33:49

13	155 bp	1070 bp	*30:01:01-30:02:12, 30:04:01-30:04:02, 30:06-30:10, 30:13-30:17, 30:19-30:20, 30:23-30:40, 30:42-30:54, 30:56-30:70N, 30:72-30:78N	
14[6]	210 bp	1070 bp	*30:13, 30:16, 30:44, 30:46	
15	245 bp	**800 bp**	*30:14L, 30:29	
16	265 bp	1070 bp	*30:15, 30:33	
17	225 bp	1070 bp	*30:01:01-30:01:07, 30:11:01-30:11:02, 30:14L-30:20, 30:23-30:24, 30:26, 30:30-30:31, 30:35-30:43, 30:48-30:49, 30:52-30:54, 30:56, 30:58-30:60, 30:62-30:63, 30:65, 30:72-30:75, 30:78N	*03:72, 11:88, 24:19, 29:37, 68:45
18	130 bp 210 bp	**800 bp**	*30:32 *30:20, 30:71	
19[5]	125 bp 160 bp 200 bp 235 bp	1070 bp	*30:70N *30:22 *30:31 *30:77	 *03:45
20	210 bp 275 bp	**800 bp**	*30:23 *30:34	 *03:04:02-03:04:03, 11:153:01
21	155 bp 185 bp	1070 bp	*30:30 *30:24	
22	180 bp 215 bp	**800 bp**	*30:36, 30:73N *30:27N	
23	155 bp	1070 bp	*30:26	
24[7]	200 bp	**800 bp**	*30:25	
25	170 bp	1070 bp	*30:40	
26[5]	80 bp	1070 bp	*30:42	
27	145 bp 195 bp	1070 bp	*30:56 *30:45	
28	270 bp	1070 bp	*30:51	*02:121, 02:425, 03:154, 23:47, 24:234, 29:06, 31:51, 68:14
29	130 bp 190 bp	1070 bp	*30:59N, 30:61, 30:74 *30:73N	*01:57N, 02:156, 02:338, 03:17:01, 03:171, 11:119:01-11:119:02, 24:92, 68:103
30[5]	80 bp	1070 bp	*30:37	
31	150 bp	1070 bp	*30:76N	
32[9]			**Negative Control**	

[1]Alleles are assigned by the presence of specific PCR product(s). However, the sizes of the specific PCR products may be helpful in the interpretation of HLA-A*30 SSP typings.
When the primers in a primer mix can give rise to HLA-specific PCR products of more than one length this is indicated if the size difference is more than 20 base pairs. Size differences of 20 base pairs or less are not given. For high resolution SSP kits, the alleles listed are specified according to amplicon length.
Nonspecific amplifications, i.e. a ladder or a smear of bands, may sometimes be seen. GC-rich primers have a higher tendency of giving rise to nonspecific amplifications than other primers.

PCR fragments longer than the control bands may sometimes be observed. Such bands should be disregarded and do not influence the interpretation of the SSP typings.

PCR fragments migrating faster than the control bands, but slower than a 400 bp fragment may be seen in some gel read-outs. Such bands can be disregarded and do not influence the interpretation of the SSP typings.

Some primers may give rise to primer oligomer artifacts. Sometimes this phenomenon is an inherit feature of the primer pair(s) of a primer mix. More often it is due to other factors such as too low amount of DNA in the PCR reactions, taking too long time in setting up the PCR reactions, working at elevated room temperature or using thermal cyclers that are not pre-heated

[2]The internal positive control primer pairs amplify segments of the human growth hormone gene. The internal positive control bands are 1070 or 800 base pairs respectively, well distribution as outlined in the table. Well number 1 contains the shorter, 800 bp, internal positive control band. The well distribution of the internal controls can help in orientation of the kit on gel photo, as well as allow for kit identification. In the presence of a specific amplification the intensity of the control band often decreases.

[3]For several HLA Class I alleles 1st and/or 4th exon(s) and beyond, as well as intron nucleotide sequences, are not available. In these instances it is not known whether some of the primers of the SSP sets are completely matched with the target sequences or not. Assumption is made that unknown sequences in these regions are conserved within allelic groups.

[4]Due to the sharing of sequence motifs between HLA-A alleles non-HLA-A*30 alleles will be amplified by primer mixes 1, 3 to 5, 7, 9 to 12, 17, 19, 20, 28 and 29.

[5]HLA-specific PCR products shorter than 125 base pairs have a lower intensity and are less sharp than longer PCR products.

[6]Primer mixes 4, 12 and 14 may give rise to nonspecific amplifications, most pronounced in primer mixes 12 and 14.

[7]Primer mix 24 may have a tendency of giving rise to primer oligomer formation.

[8]Primer mixes 3 and 8 may give rise to a lower yield of HLA-specific PCR product than the other A*30 primer mixes.

[9]Primer mix 32 contains a negative control, which will amplify more than 95% of HLA amplicons as well as the amplicons generated by control primer pairs. PCR product sizes range from 75 to 200 base pairs. The PCR product generated by the control primer pair is 430 base pairs.

'w', might be weakly amplified.

101.429-12 – including *Taq* polymerase, IFU-01
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PRIMER SPECIFICATION

Well No.	1	2	3	4	5	6	7	8	9	10	11	12
Length of spec. PCR product	165	205	210	155	150 245	80 185	200	85	180	150	215	190
Length of int.	800	800	800	1070	1070	1070	1070	800	1070	1070	800	1070
pos. control[1]												
5'-primer(s)[2]	414 5' -gAA 3'	363 5' -ATA 3'	363 5' -ATA 3'	123 5' -AgT 3'	123 5' -AgT 3' 414 5' -gAA 3'	123 5' -AgT 3'	78 5' -TCT 3' 362 5' -ggT 3'	367 5' -TgC 3'	98 5' -CTC 3'	127 5' -ggg 3'	98 5' -CTC 3'	363 5' -ATA 3'
3'-primer(s)[3]	538 5' -CAA 3'	526 5' -CCA 3'	526 5' -CCg 3' 535 5' -CTA 3' 538 5' -CAg 3'	238 5' -CCC 3'	325 5' -gTg 3' 524 5' -CAT 3'	163 5' -CgC 3' 265 5' -CCC 3'	238 5' -CCT 3' 526 5' -CCg 3'	411 5' -TCA 3'	238 5' -CCT 3'	238 5' -CCT 3'	270 5' -ACA 3'	505 5' -gCT 3' 524 5' -CAC 3'
Well No.	1	2	3	4	5	6	7	8	9	10	11	12

Well No.	13	14	15	16	17	18	19	20	21	22	23	24
Length of spec. PCR product	155	210	245	265	225	130 210	125 160 200 235	210 275	155 185	180 215	155	105 200
Length of int.	1070	1070	800	1070	1070	800	1070	800	1070	800	1070	800
pos. control[1]												
5'-primer(s)[2]	123 5' -AgT 3'	123 5' -AgT 3'	363 5' -ATA 3'	363 5' -ATA 3' 784 5' -ggA 3'	98 5' -CTC 3'	123 5' -AgT 3'	123 5' -AgT 3' 650 5' -CCC 3'	363 5' -ATA 3'	410 5' -gTg 3' 634 5' -CAg 3'	363 5' -ATA 3'	413 5' -CCg 3'	123 5' -AgT 3'
3'-primer(s)[3]	238 5' -CCT 3'	292 5' -gTg 3'	563 5' -Cgg 3' 572 5' -gCg 3'	586 5' -CAC 3' 899 5' -ACA 3'	282 5' -gAC 3'	212 5' -gCC 3' 289 5' -AgC 3' 294 5' -CgT 3'	206 5' -.CA 3' 240 5' -ggA 3' 281 5' -ACC 3' 845 5' -AgT 3'	530 5' -CCT 3' 595 5' -CCT 3'	526 5' -CCA 3' 777 5' -gCA 3'	494 5' -TCC 3' 513 5' -TCC 3' 535 5' -CTA 3'	526 5' -CCA 3'	187 5' -g.C 3' 282 5' -gAC 3'
Well No.	13	14	15	16	17	18	19	20	21	22	23	24

CE 0088

101.429-12 – including *Taq* polymerase, IFU-01
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Well No.	25	26	27	28	29	30	31
Length of spec.	170	80	145	270	130	80	150
PCR product			195		190		
Length of int.	1070	1070	1070	1070	1070	1070	1070
pos. control[1]							
5'-primer(s)[2]	397	363	176	385	363	485	127
	5' -gCC 3'	5' -ATA 3'	5' -gCA 3'	5' -ggT 3'	5' -ATA 3'	5' -CAC 3'	5' -gAT 3'
			414				
			5' -gAA 3'				
3'-primer(s)[3]	526	403	282	616	453	526	238
	5' -CCA 3'	5' -gCT 3'	5' -gAC 3'	5' -CgC 3'	5' -TCT 3'	5' -CCA 3'	5' -CCT 3'
			570		454		
			5' -CAg 3'		5' -CTA 3'		
					513		
					5' -TCC 3'		
Well No.	25	26	27	28	29	30	31

[1]The internal positive control primer pairs amplify segments of the human growth hormone gene. The internal positive control bands are 1070 or 800 base pairs respectively, well distribution as outlined in the table. Well number 1 contains the shorter, 800 bp, internal positive control band. The well distribution of the internal controls can help in orientation of the kit on gel photo, as well as allow for kit identification. In the presence of a specific amplification the intensity of the control band often decreases.

[2]The nucleotide position matching the specificity-determining 3'-end of the primer is given. Nucleotide numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

[3]The nucleotide position matching the specificity-determining 3'-end of the primer is given in the anti-sense direction. Nucleotide numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

101.429-12 – including *Taq* polymerase, IFU-01
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CELL LINE VALIDATION SHEET

HLA-A*30 SSP subtyping kit[2]

					Well													
					1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16
				Lot No.:	201206201	201206202	201206203	201206204	201206205	201206206	201206207	201206208	201206209	201206210	201206211	201206212	201206213	201206214	201206215	201206216
	IHWC	cell line[1]	A*	A*																
1	9001	SA	*24:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
2	9280	LK707	*02:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
3	9011	E4181324	*01:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
4	9275	GU373	*30:01		+	+	-	-	-	-	-	-	+	-	+	-	+	-	-	-
5	9009	KAS011	*01:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
6	9353	SM	*02:01	*26:03	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
7	9020	QBL	*26:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
8	9025	DEU	*31:01		-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-
9	9026	YAR	*26:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
10	9107	LKT3	*24:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
11	9051	PITOUT	*29:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
12	9052	DBB	*02:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
13	9004	JESTHOM	*02:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
14	9071	OLGA	*31:01		-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-
15	9075	DKB	*24:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
16	9037	SWEIG007	*29:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
17	9282	CTM3953540	*03:01	*80:01	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
18	9257	32367	*33:03	*74:01	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
19	9038	BM16	*02:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
20	9059	SLE005	*02:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
21	9064	AMALA	*02:17		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
22	9056	KOSE	*02:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
23	9124	IHL	*02:01	*34:01	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
24	9035	JBUSH	*32:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
25	9049	IBW9	*33:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
26	9285	WT49	*02:05		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
27	9191	CH1007	*24:10	*29:01	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
28	9320	BEL5GB	*02:01	*29:02	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
29	9050	MOU	*29:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
30	9021	RSH	*30:01	*68:02	+	+	-	-	-	-	-	-	+	-	+	-	+	-	-	-
31	9019	DUCAF	*30:02		+	-	+	-	-	-	-	-	+	-	+	-	+	-	-	-
32	9297	HAG	*02:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
33	9098	MT14B	*31:01		-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-
34	9104	DHIF	*31:01		-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-
35	9302	SSTO	*32:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
36	9024	KT17	*02:06	*11:01	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
37	9065	HHKB	*03:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
38	9099	LZL	*02:17		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
39	9315	CML	*01:01	*03:01	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
40	9134	WHONP199	*02:07	*30:01	+	+	-	-	-	-	-	-	+	-	+	-	+	-	-	-
41	9055	H0301	*03:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
42	9066	TAB089	*02:07		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
43	9076	T7526	*02:06	*02:07	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
44	9057	TEM	*66:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
45	9239	SHJO	*23:01	*24:02	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
46	9013	SCHU	*03:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
47	9045	TUBO	*02:16	*03:01	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
48	9303	TER-ND	*02:01	*11:01	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-

101.429-12 – including *Taq* polymerase, IFU-01
101.429-12u – without *Taq* polymerase, IFU-02

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CELL LINE VALIDATION SHEET

HLA-A*30 SSP subtyping kit[2]

					Well														
					17	18	19	20	21	22	23	24	25	26	27	28	29	30	31
				Lot No.:	201206217	201430918	201430919	201206220	201206221	201430922	201430923	201430924	201430925	201206226	201430927	201206228	201430929	201206230	201430931
	IHWC cell line[1]		A*	A*															
1	9001	SA	*24:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
2	9280	LK707	*02:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
3	9011	E4181324	*01:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
4	9275	GU373	*30:01		+	-	-	-	-	-	-	-	-	-	-	-	-	-	-
5	9009	KAS011	*01:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
6	9353	SM	*02:01	*26:03	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
7	9020	QBL	*26:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
8	9025	DEU	*31:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
9	9026	YAR	*26:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
10	9107	LKT3	*24:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
11	9051	PITOUT	*29:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
12	9052	DBB	*02:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
13	9004	JESTHOM	*02:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
14	9071	OLGA	*31:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
15	9075	DKB	*24:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
16	9037	SWEIG007	*29:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
17	9282	CTM3953540	*03:01	*80:01	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
18	9257	32367	*33:03	*74:01	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
19	9038	BM16	*02:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
20	9059	SLE005	*02:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
21	9064	AMALA	*02:17		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
22	9056	KOSE	*02:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
23	9124	IHL	*02:01	*34:01	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
24	9035	JBUSH	*32:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
25	9049	IBW9	*33:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
26	9285	WT49	*02:05		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
27	9191	CH1007	*24:10	*29:01	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
28	9320	BEL5GB	*02:01	*29:02	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
29	9050	MOU	*29:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
30	9021	RSH	*30:01	*68:02	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-
31	9019	DUCAF	*30:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
32	9297	HAG	*02:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
33	9098	MT14B	*31:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
34	9104	DHIF	*31:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
35	9302	SSTO	*32:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
36	9024	KT17	*02:06	*11:01	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
37	9065	HHKB	*03:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
38	9099	LZL	*02:17		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
39	9315	CML	*01:01	*03:01	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
40	9134	WHONP199	*02:07	*30:01	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-
41	9055	H0301	*03:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
42	9066	TAB089	*02:07		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
43	9076	T7526	*02:06	*02:07	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
44	9057	TEM	*66:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
45	9239	SHJO	*23:01	*24:02	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
46	9013	SCHU	*03:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
47	9045	TUBO	*02:16	*03:01	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
48	9303	TER-ND	*02:01	*11:01	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-

[1]The provided cell line HLA specificities are retrieved from the http://www.ihwg.org/hla web site. The specificity of an individual cell line may thus be subject to change.

101.429-12 – including *Taq* polymerase, IFU-01
101.429-12u – without *Taq* polymerase, IFU-02

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Lot No.: 38V **Lot-specific Information** **www.olerup-ssp.com**

[2]The specificity of each primer solution in the kit has been tested against 48 well characterized cell line DNAs and where applicable, additional cell line DNAs.
No DNAs carrying the alleles to be amplified by primer solutions 6 to 7, 14 to 16 and 18 to 31 were available. The specificities of the primers in primer 6, 7, 14 to 16, 19, 23, 24, 27, 29 and 30 were tested by separately adding one additional 5'-primer, respectively one additional 3'-primer. In primer solutions 18, 20, 22, 26 and 28 it was only possible to test the 5'-primer, the 3'-primers were not possible to test. In primer solution 21, 25 and 31 it was only possible to test the 3'-primers, the 5'-primers were not possible to test.
In primer solutions 7 and 16 one 5'-primer was not possible to test, and in primer solutions 2, 3, 5, 6, 12, 15, 16, 19, 24, 27 and 29 one or two 3'-primers were not possible to test.
In addition, one 5'-primer in primer solution 5 was tested by adding an additional 3'-primer.

101.429-12 – including *Taq* polymerase, IFU-01
101.429-12u – without *Taq* polymerase, IFU-02

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101.429-12 – including *Taq* polymerase, IFU-01
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